

Preparation of *N*-^tBoc L-glutathione dimethyl and di-*tert*-butyl esters: Versatile synthetic building blocks

J. R. Falck,^{a,*} Bhavani Sangras^a and Jorge H. Capdevila^b

^aDepartments of Biochemistry and Pharmacology, University of Texas Southwestern Medical Center, Dallas, TX 75390-9038, USA

^bDepartments of Medicine and Biochemistry, Vanderbilt University School of Medicine, Nashville, TN 37232, USA

Received 16 September 2006; accepted 11 October 2006

Available online 13 October 2006

Abstract—The title L-glutathione derivatives, containing acid- and base-labile esters, respectively, were obtained in good overall yields. *N*-^tBoc L-glutathione dimethyl ester was prepared via Fischer esterification of L-glutathione disulfide (GSSG) using HCl in dry methanol, protection of the amine with ^tBoc₂O, and tributylphosphine cleavage of the disulfide in wet isopropanol. Alternatively, Fischer esterification and ^tBoc-protection of L-glutathione (GSH) also furnished *N*-^tBoc glutathione dimethyl ester accompanied by a small amount of *S*-^tBoc that was removed chromatographically. The di-*tert*-butyl ester was obtained by *S*-palmitoylation of GSH in TFA as solvent, *N*-^tBoc-protection, esterification using ^tBuOH mediated by diisopropylcarbodiimide/copper(I) chloride, and saponification of the thioester. These L-glutathione derivatives are versatile synthetic building blocks for the preparation of *S*-glutathione adducts.

© 2006 Elsevier Ltd. All rights reserved.

1. Introduction

The tripeptide L-glutathione (L- γ -glutamyl-L-cysteinylglycine; GSH) (**4**) is a common constituent in most animal cells¹ and many bacteria.² It participates in a variety of critical physiologic functions, *inter alia*, signal transduction,³ immunity,⁴ maintenance of cellular osmolality,⁵ defense against reactive oxygen species (ROS) and free radicals,⁶ and protein folding.⁷ Of particular interest is the bioactivation⁸ or inactivation of xenobiotics, drugs, and endogenous substrates by GSH-*S*-conjugation mediated by a widely distributed family of GSH-*S*-transferases.⁹ The latter category of substrates includes metabolites of the cyclooxygenase, lipoxygenase, and cytochrome P450 branches of the arachidonate cascade.¹⁰ As part of our longstanding interest in the structure elucidation and total synthesis of GSH-*S*-conjugates of eicosanoids,¹¹ we required *N*-protected L-glutathione derivatives bearing orthogonally protected esters as synthetic intermediates. Herein, we report reliable, multi-gram preparations of the base-labile building block *N*-^tBoc L-glutathione dimethyl ester¹² (**3**) and its acid-labile di-*tert*-butyl ester analog (**9**).

2. Results and discussion

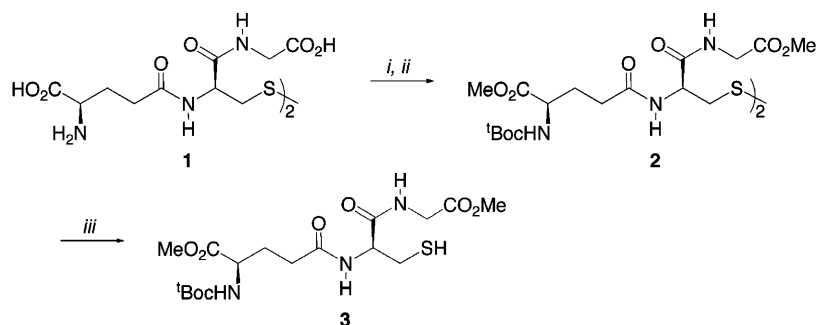
The synthesis of **3** began by dissolving commercial L-glutathione disulfide (**1**) in MeOH and saturating with dry HCl gas (Scheme 1).¹³ After several days, all volatiles were removed in vacuo and, typically, the crude tetramethyl ester dihydrochloride salt¹³ was directly *N*-carbamoylated using ^tBoc-anhydride in the presence of NaHCO₃ to give **2**.¹⁴ Disulfide cleavage¹⁵ using *n*-Bu₃P proceeded smoothly and furnished *N*-^tBoc L-glutathione dimethyl ester¹² (**3**) as a white, crystalline solid in 54% overall yield.

Alternatively, Fischer esterification of **4** as conducted above and subsequent *N*-carbamoylation of the resultant dimethyl ester hydrochloride salt¹⁶ **5** led to **3** (Scheme 2).¹⁷ The somewhat superior yield of the route in Scheme 2 (64% overall) versus that in Scheme 1 (54% overall) is counterbalanced by the need to remove chromatographically a small amount of *N,S*-di-^tBoc dimethyl ester by-product.

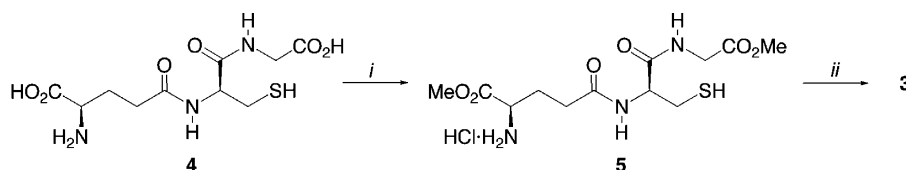
Due in large part to its poor solubility in even polar organic solvents, for example, DMF, DMSO, dioxane, and THF, the conversion of **4** to its di-*tert*-butyl ester proved problematic. Little or no esterification was noted using a variety of chemical and enzymatic procedures: isobutylene/H₂SO₄ or CH₃SO₃H in dioxane,¹⁸ ^tBuOAc/

Keywords: Amino acids and derivatives; Thioesters; Protecting groups; Peptides.

* Corresponding author. Tel.: +1 214 648 2406; fax: +1 214 648 6455; e-mail: j.falck@UTSouthwestern.edu



Scheme 1. Reagents and conditions: (i) HCl, MeOH, 0 °C, 80 h; (ii) t Boc₂O, NaHCO₃, THF/H₂O, 23 °C, 29 h, 75% from **1**; (iii) Bu₃P, *n*-PrOH/H₂O (2:1), 23 °C, 4 h, 72%.



Scheme 2. Reagents and conditions: (i) HCl, MeOH, 0 °C, 80 h, 91%; (ii) t Boc₂O, NaHCO₃, THF/H₂O, 23 °C, 15 h, 70%.

HClO₄,¹⁹ t Boc₂O/DMAP/ t BuOH,²⁰ t BuBr/K₂CO₃/(PhCH₂)Et₃NCl in DMF or THF,²¹ t BuOC(O)F/Et₃N/DMAP in t BuOH under reflux,²² Me₂NC(O t Bu)₂H,²³ t BuOH/EDCI/DMAP,²⁴ t BuOH/CDI/DBU,²⁵ transesterification of **5** with t BuOH/H₂SO₄, t BuOH/amano AK lipase, and t BuOH/pig liver esterase.²⁶ To improve its solubility characteristics as well as obviate the inherent nucleophilicity of the thiol, **4** was dissolved in a minimum of TFA and *S*-acylated with acid chlorides of varying chain lengths.^{27,28} The palmitoyl thioester **6** displayed satisfactory behavior and could be isolated as the solid trifluoroacetate salt (Scheme 3). No *N*-palmitoylation under the acidic conditions was noted.²⁹ Sequential *N*-carbamoylation and esterification of **6** with t BuOH afforded *N*-Boc **7** and *N*-Boc di-*tert*-butyl ester **8**, respectively. The latter esterification, however, could only be effected in satisfactory yield using *N,N'*-diisopropylcarbodiimide in the presence of catalytic CuCl according to Zhu et al.³⁰ Methanolysis³¹ of **8** with NaOMe/MeOH gave rise to **9** without event.

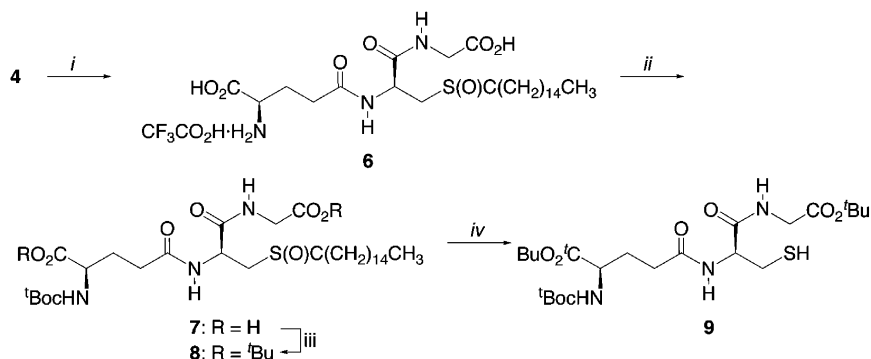
3. Conclusions

Herein, we provide convenient and efficient preparations of the dimethyl and di-*tert*-butyl esters of *N*- t Boc L-glutathione. Since these synthetic building blocks can be orthogonally deprotected under basic and acidic conditions, respectively, we anticipate they will find utility in the preparation of *S*-glutathione conjugates, peptide synthesis, and library development.

4. Experimental

4.1. General procedures

¹H and ¹³C spectra were recorded in CDCl₃ unless otherwise specified using tetramethylsilane as internal reference. The Michigan State University Mass Spectroscopy Facility provided high-resolution mass spectra. All reactions were maintained under an argon



Scheme 3. Reagents and conditions: (i) H₃C(CH₂)₁₄C(O)Cl, F₃CCO₂H, 23 °C, 20 min, then 40 °C, 40 min, 91%; (ii) O(CO t Bu)₂, NaHCO₃, THF/H₂O, 23 °C, 15 h, 64%; (iii) t BuOH, (PrHN=)₂C, CuCl, 23 °C, 12 h, then add **7** in CH₂Cl₂, 40 °C, 48 h, 70%; (iv) NaOMe, MeOH, 1 h, 23 °C, 71%.

atmosphere. Anhydrous solvents were freshly distilled from sodium benzophenone ketyl, except for CH_2Cl_2 , which was distilled from CaH_2 . Extracts were dried over anhydrous Na_2SO_4 and filtered prior to removal of all volatiles under reduced pressure.

4.2. Chemistry

4.2.1. Bis-*N*-^tBoc L-glutathione tetramethyl ester disulfide (2). Dry HCl gas was bubbled through a 0 °C solution of L-glutathione disulfide (**1**) (6.0 g, 9.8 mmol) in anhydrous MeOH (500 mL) until 48 g of HCl was absorbed. Stirring was continued at 0 °C for 80 h, then all volatiles were removed in vacuo. The residue was further dried using a mechanical vacuum pump for 24 h to give the corresponding tetramethyl ester dihydrochloride salt as a colorless gum that was used directly in the next step.

A solution of the above tetramethyl ester salt (6.0 g), NaHCO_3 (3.3 g, 39.2 mmol, 4 equiv), and ^tBoc₂O (5.16 g, 23 mmol, 2.4 equiv) in THF/H₂O (2.4:1, 150 mL) was stirred at room temperature. After 29 h, the pH was adjusted to 3 with concd HCl, and the solution was extracted with EtOAc (5 × 10 mL). The combined extracts were concentrated in vacuo and the residue purified by SiO₂ column chromatography to give **2** (4.0 g, 75%) as a white solid, mp 135 °C. TLC: 10% MeOH/ CH_2Cl_2 , $R_f \sim 0.35$; $[\alpha]_D^{23} + 30.5$ (c 2.25, CHCl_3); ¹H NMR (400 MHz) δ 8.46–8.58 (m, 1H), 6.72 (d, 1H, $J = 8.6$ Hz), 5.51–5.56 (m, 1H), 5.32 (d, 1H, $J = 8.2$ Hz), 4.34–4.44 (m, 1H), 4.15 (dd, 1H, $J = 5.8$, 18 Hz), 4.09 (dd, 1H, $J = 4.9$, 18 Hz), 3.76 (s, 3H), 3.74 (s, 3H), 3.06 (dd, 1H, $J = 4$, 15 Hz), 2.84–2.97 (m, 1H), 2.32–2.44 (m, 2H), 2.14–2.26 (m, 1H), 1.95–2.20 (m, 1H), 1.43 (s, 9H); ¹³C NMR (100 MHz) δ 173.2, 172.8, 171.2, 170.1, 155.9, 80.2, 53.2, 53.1, 52.5, 46.1, 41.5, 32.4, 28.5.

4.2.2. *N*-^tBoc L-glutathione dimethyl ester (3) from 2. Bu₃P (500 μL , 617 mg, 3.05 mmol) was added to a solution of disulfide **2** (1.6 g, 1.84 mmol) in *n*-PrOH/H₂O (2:1, 6 mL, degassed with argon) under argon atmosphere. After stirring for 4 h, the *n*-PrOH was removed and the aqueous solution was extracted with CH_2Cl_2 (4 × 40 mL). The combined extracts were washed with water, dried, and concentrated. The residue was purified by SiO₂ column chromatography to give **3** (1.16 g, 72%) as a white, crystalline solid, mp 94 °C. TLC: 10% MeOH/ CH_2Cl_2 , $R_f \sim 0.38$; $[\alpha]_D^{23} - 6.8$ (c 1.3, CHCl_3); ¹H NMR (400 MHz) δ 6.97 (t, 1H, $J = 5.5$ Hz), 6.83 (d, 1H, $J = 7$ Hz), 5.29 (d, 1H, $J = 8.5$ Hz), 4.68 (ddd, 1H, $J = 8.5$, 6.0, 4.6 Hz), 4.30–4.40 (m, 1H), 4.08 (dd, 1H, $J = 5.8$, 18 Hz), 4.00 (dd, 1H, $J = 18$, 5.2 Hz), 3.76 (s, 3H), 3.75 (s, 3H), 3.14 (ddd, 1H, $J = 14$, 7.9, 4.6 Hz), 2.74–2.84 (m, 1H), 2.38 (t, 2H, $J = 6.5$ Hz), 2.20–2.26 (m, 1H), 1.90–1.99 (m, 1H), 1.83 (dd, 1H, $J = 8$, 11 Hz), 1.43 (s, 9H); ¹³C NMR (100 MHz) δ 173.1, 172.8, 170.4, 170.2, 86.04, 80.3, 54.1, 52.7, 52.5, 41.4, 32.1, 28.5, 28.3, 26.4.

4.2.3. L-Glutathione dimethyl ester hydrochloride (5). L-Glutathione (**4**) (2.0 g, 6.51 mmol) was esterified in acidified MeOH as described for **2** to give **5** (2.2 g, 91%) as a free flowing, white solid, mp 98 °C. TLC:

10% MeOH/ CH_2Cl_2 , $R_f \sim 0.25$; $[\alpha]_D^{23} - 26$ (c 1.1, EtOH); ¹H NMR (400 MHz, CD_3OD) δ 4.58 (t, 1H, $J = 6.4$ Hz), 4.01 (d, 1H, $J = 5.5$ Hz), 3.78 (s, 3H), 3.76 (s, 3H), 3.62 (t, 1H, $J = 6.8$ Hz), 3.32–3.37 (m, 1H), 2.95 (dd, 1H, $J = 6.7$, 14.2 Hz), 2.87 (dd, 1H, $J = 7.2$, 13.9 Hz), 2.43–2.51 (m, 2H), 2.06–2.15 (m, 1H), 1.93–2.04 (m, 2H); ¹³C NMR (100 MHz, CD_3OD) δ 176.3, 175.3, 173.0, 171.7, 57.0, 54.7, 52.9, 52.81, 42.0, 32.9, 30.7, 27.1.

4.2.4. *N*-^tBoc L-glutathione dimethyl ester (3) from 5. A solution of **5** (2.2 g, 6.6 mmol), NaHCO_3 (1.2 g, 14.3 mmol, 2.2 equiv), and ^tBoc₂O (1.4 g, 6.6 mmol, 1.0 equiv) in THF/H₂O (1:2.4, 25 mL) was stirred at room temperature. After 15 h, the solution was extracted with EtOAc (4 × 50 mL). The combined extracts were concentrated in vacuo and the residue was purified by SiO₂ column chromatography to give **3** (1.8 g, 70%) as described above and the by-product *S*-^tBoc, *N*-^tBoc dimethyl ester (0.39 g, 11%) as a white solid, mp 91 °C; lit.¹⁷ mp 92 °C. TLC of *S*-^tBoc, *N*-^tBoc dimethyl ester: 5% MeOH/ CH_2Cl_2 , $R_f \sim 0.25$; $[\alpha]_D^{23} - 32$ (c 1.0, EtOH); lit.¹⁷ $[\alpha]_D^{23} - 37.2$ (c 1.0, EtOH); ¹H NMR (400 MHz) δ 7.29 (t, 1H, $J = 5.2$ Hz), 6.93 (d, 1H, $J = 7.2$ Hz), 5.44 (d, 1H, $J = 7.9$ Hz), 4.63–4.69 (m, 1H), 4.25 (d, 1H, $J = 4.7$ Hz), 4.02 (dd, 1H, $J = 5.7$, 17.9 Hz), 3.95 (dd, 1H, $J = 4.7$, 18 Hz), 3.69 (s, 3H), 3.68 (s, 3H), 3.22 (dd, 1H, $J = 4.8$, 15 Hz), 3.13 (dd, 1H, $J = 7.0$, 14.6 Hz), 2.29 (t, 2H, $J = 7.2$ Hz), 2.08–2.14 (m, 1H), 1.92–2.00 (m, 1H), 1.43 (s, 9H), 1.38 (s, 9H); ¹³C NMR (100 MHz) δ 173.0, 172.7, 170.5, 170.2, 169.9, 155.8, 85.8, 80.2, 53.7, 53.1, 53.0, 52.6, 52.5, 41.4, 32.3, 32.2, 28.4, 28.2.

4.2.5. *S*-Palmitoyl L-glutathione trifluoroacetate (6). Palmitoyl chloride (2.4 mL, 7.8 mmol, 2.4 equiv) was added dropwise to a solution of L-glutathione (**4**) (1.0 g, 3.25 mmol) in trifluoroacetic acid (11 mL) under an argon atmosphere. After stirring at room temperature for 20 min and at 40 °C for 30 min, the reaction was quenched by the addition of water (0.25 mL, 14 mmol, 4.3 equiv) and the reaction mixture was stirred for an additional 1 h at 40 °C. The trifluoroacetic acid was removed in vacuo, ethyl acetate (20 mL) was added, the mixture was cooled to 10 °C, and the precipitated trifluoroacetate salt of **6** (1.6 g, 91%) was collected by filtration, mp 179–181 °C. TLC: MeOH/H₂O (2:1), $R_f \sim 0.67$; $[\alpha]_D^{23} - 15$ (c 0.66, EtOH/DMSO (2:1)); ¹H NMR (400 MHz, CD_3SOCD_3) δ 8.24–8.37 (m, 1H), 4.36–4.45 (m, 1H), 3.60–3.86 (m, 1H), 3.24–3.38 (m, 1H), 2.92–2.99 (m, 1H), 2.84–2.97 (m, 1H), 2.25–2.56 (m, 2H), 2.16 (t, 1H, $J = 7.3$ Hz), 1.10–1.38 (m, 18H), 0.83 (t, 3H, $J = 6.8$ Hz); ESMS m/z 546 ($\text{M}^+ + 1$); HRMS (FAB-Cl, NBA) Calcd for $\text{C}_{26}\text{H}_{47}\text{N}_3\text{O}_7\text{S}$ [$\text{M} + \text{H}$]⁺ 546.3214, found 546.3213.

4.2.6. *N*-^tBoc *S*-palmitoyl L-glutathione (7). Sodium bicarbonate (984 mg, 11.72 mmol, 4 equiv) and di-*tert*-butyldicarbonate (767 mg, 3.5 mmol, 1.2 equiv) were added sequentially to a solution of **6** (1.6 g, 2.93 mmol) in THF/H₂O (1:2.4, 10 mL) under an argon atmosphere. After stirring overnight, the pH was adjusted to 4 with 6 N HCl and the reaction mixture was extracted with EtOAc (4 × 5 mL). The

combined organic extracts were dried, concentrated in vacuo, and the solid white residue was azeotroped with anhydrous benzene (20 mL) and dried under high vacuum for 1 h to yield **7** (1.2 g, 64%) as a free flowing solid, mp 104 °C, sufficiently pure to be used directly in the next step. TLC: 30% MeOH/EtOAc, $R_f \sim 0.25$; $[\alpha]_D^{23} - 8$ (c 1.7, CHCl_3); ^1H NMR (400 MHz) δ 7.50–8.20 (m, 2H), 6.47 (br s, 1H), 5.65 (br s, 1H), 4.71 (d, 1H, $J = 5.8$ Hz), 4.13–4.35 (m, 2H), 3.72–3.88 (m, 2H), 3.18–3.36 (m, 2H), 2.55 (t, 2H, $J = 7.6$ Hz), 2.35 (t, 2H, $J = 7.3$ Hz), 1.45 (s, 9H), 1.25 (s, 26H), 0.88 (t, 3H, $J = 6.7$ Hz); ^{13}C NMR (75 MHz) δ 200.1, 179.4, 175.4, 174.2, 172.8, 171.6, 156.2, 85.3, 82.2, 80.6, 53.0, 44.2, 41.6, 34.3, 32.1, 29.9, 29.7, 29.6, 29.5, 29.3, 29.2, 28.5, 27.6, 27.8, 25.0, 22.9, 14.3; ESMS m/z 668 ($\text{M}^+ + 23$); HRMS (FAB-Cl, NBA) Calcd for $\text{C}_{31}\text{H}_{55}\text{N}_3\text{O}_9\text{S}$ $[\text{M} + \text{H}]^+$ 646.3737, found 646.3737.

4.2.7. *N*'-Boc *S*-palmitoyl L-glutathione di-*tert*-butyl ester (8). *tert*-Butanol (2.18 mL, 23 mmol, 10.6 equiv) and *N,N'*-diisopropylcarbodiimide (3.2 mL, 20.8 mmol, 9.6 equiv) were stirred overnight in the presence of a catalytic amount of CuCl (25 mg) under an argon atmosphere. The resulting *O-tert*-butyl *N,N'*-diisopropylisourea solution was diluted with dry dichloromethane (10 mL) followed by the addition of **7** (1.4 g, 2.16 mmol) and then the reaction mixture was heated under reflux. After 2 days, the reaction mixture was filtered through Celite® and the filter cake was washed with dichloromethane (150 mL). The filtrate was concentrated in vacuo and the residue purified by column chromatography on silica gel impregnated with triethylamine (2 mL $\text{Et}_3\text{N}/100$ g SiO_2) using 30% EtOAc/hexanes as eluant to give **8** (1.15 g, 70%) as a thick syrup. TLC: 5% MeOH/ CH_2Cl_2 , $R_f \sim 0.66$; $[\alpha]_D^{23} - 12.5$ (c 2.06, CHCl_3); ^1H NMR (300 MHz) δ 6.96 (t, 1H, $J = 5.2$ Hz), 6.77 (d, 1H, $J = 7.0$ Hz), 5.24 (d, 1H, $J = 7.6$ Hz), 4.60 (dt, 1H, $J = 4.8, 7.9$ Hz), 4.16–4.22 (m, 1H), 3.90 (dd, 1H, $J = 5.5, 12.6$ Hz), 3.34 (dd, 1H, $J = 4.6, 14.3$ Hz), 3.25 (dd, 1H, $J = 7.94, 14.3$ Hz), 2.56 (t, 2H, $J = 7.3$ Hz), 2.12–2.38 (m, 2H), 1.63 (t, 2H, $J = 7.1$ Hz), 1.46 (s, 9H), 1.45 (s, 9H), 1.43 (s, 9H), 1.24 (br s, 26H), 0.87 (t, 3H, $J = 7.0$ Hz); ^{13}C NMR (75 MHz) δ 200.5, 172.8, 171.5, 170.1, 168.6, 155.9, 82.3, 80.0, 53.6, 44.1, 42.2, 32.4, 32.0, 30.4, 29.78, 29.77, 29.75, 29.71, 29.54, 29.47, 29.3, 29.1, 28.8, 28.5, 28.2, 28.1, 25.7, 22.8, 14.3; ESMS m/z 780 ($\text{M}^+ + 23$); HRMS (FAB-Cl, NBA) Calcd for $\text{C}_{39}\text{H}_{71}\text{N}_3\text{O}_9\text{S}$ $[\text{M} + \text{H}]^+$ 758.4997, found 758.4989.

4.2.8. *N*'-Boc L-glutathione di-*tert*-butyl ester (9). A freshly prepared 0.1 N solution of NaOMe in MeOH (14 mL, 1.45 mmol) was added to a stirring solution of **8** (1.0 g, 1.32 mmol) in methanol (14 mL) under an argon atmosphere. After 1 h, the reaction mixture was cooled to 5 °C and acidified to pH 5 using 5% acetic acid in ether. All volatiles were removed in vacuo and the residue was purified by column chromatography using 50% EtOAc/hexanes as eluant to furnish **9** (480 mg, 71%), mp 41.5 °C. TLC: 50% EtOAc/hexanes, $R_f \sim 0.26$; $[\alpha]_D^{23} - 7.6$ (c 2.0, CHCl_3);

^1H NMR (400 MHz) δ 6.93 (d, 1H, $J = 6.4$ Hz), 6.80–6.85 (m, 1H), 5.26 (d, 2H, $J = 7.9$ Hz), 4.64–4.72 (m, 1H), 4.16–4.28 (m, 1H), 3.96 (dd, 1H, $J = 5.5, 13$ Hz), 3.87 (dd, 1H, $J = 5.4, 11.7$ Hz), 3.06–3.18 (m, 2H), 2.78–2.86 (m, 1H), 2.36 (t, 2H, $J = 7.4$ Hz), 2.18–2.26 (m, 1H), 1.82 (dd, 1H, $J = 7.9, 9.8$ Hz), 1.47 (s, 18H), 1.45 (s, 9H); ^{13}C NMR (75 MHz) δ 172.6, 171.6, 170.1, 168.7, 156.0, 82.4, 80.1, 54.6, 53.5, 42.2, 32.5, 29.1, 28.4, 28.14, 28.09, 26.7; ESMS m/z 542 ($\text{M}^+ + 23$); HRMS (FAB-Cl, NBA) Calcd for $\text{C}_{23}\text{H}_{41}\text{N}_3\text{O}_8\text{S}$ $[\text{M} + \text{H}]^+$ 520.2693, found 520.2692.

Acknowledgments

Financial support was provided by the Robert A. Welch Foundation and NIH (GM31278, DK38226, GM37922).

References and notes

- Reid, M.; Jahoor, F. *Curr. Opin. Clin. Nutr. Metab. Care* **2000**, *3*, 385–390.
- Masip, L.; Veeravalli, K.; Georgiou, G. *Antioxid. Redox Signal.* **2006**, *8*, 753–762.
- Reid, M.; Jahoor, F. *Curr. Opin. Clin. Nutr. Metab. Care* **2001**, *4*, 65–71.
- Droge, W.; Breitkreutz, R. *Proc. Nutr. Soc.* **2000**, *59*, 595–600.
- Lorenson, M. Y.; Jacobs, L. S. *Endocrinology* **1987**, *120*, 365–372.
- Cnubben, N. H. P.; Rietjens, I. M. C. M.; Wortelboer, H.; van Zanden, J.; van Bladeren, P. J. *Environ. Toxicol. Pharm.* **2001**, *10*, 141–152.
- Shackelford, R. E.; Heinloth, A. N.; Heard, S. C.; Paules, R. S. *Antioxid. Redox Signal.* **2005**, *7*, 940–950.
- (a) Vamvakas, S.; Anders, M. W. *Adv. Exp. Med. Biol.* **1991**, *283*, 13–24; (b) Lauterburg, B. H. *Prog. Pharm. Clin. Pharm.* **1991**, *8*, 201–213.
- Ingelman-Sundberg, M. *Chem. Biol. Interact.* **2001**, *133*, 84–86.
- Review Murphy, R. C.; Zarini, S. *Prostaglandins Other Lipid Mediat.* **2002**, *68–69*, 471–482.
- Spearman, M. E.; Prough, R. A.; Estabrook, R. W.; Falck, J. R.; Manna, S.; Leibman, K. C.; Murphy, R. C.; Capdevila, J. *Arch. Biochem. Biophys.* **1985**, *242*, 225–230.
- For a multi-step total synthesis of **3** from L-cysteine and an unsuccessful attempt directly from L-glutathione, see (a) Threadgill, M. D.; Gledhill, A. P. *J. Org. Chem.* **1989**, *54*, 2940–2949; Using L-glutathione see (b) Crich, D.; Krishnamurthy, V.; Hutton, T. K. *J. Am. Chem. Soc.* **2006**, *128*, 2544–2545.
- Su, D.; Ren, X.; You, D.; Li, D.; Mu, Y.; Yan, G.; Zhang, Y.; Luo, Y.; Xue, Y.; Shen, J.; Liu, Z.; Luo, G. *Arch. Biochem. Biophys.* **2001**, *395*, 177–184.
- Arisawa, M.; Ono, T.; Yamaguchi, M. *Tetrahedron Lett.* **2005**, *46*, 5669–5671.
- Kedrowski, B. L.; Heathcock, C. H. *Heterocycles* **2002**, *58*, 601–634.
- Anderson, M. E.; Powrie, F.; Puri, R. N.; Meister, A. *Arch. Biochem. Biophys.* **1985**, *239*, 538–548.
- Muraki, M.; Mizoguchi, T. *Chem. Pharm. Bull.* **1971**, *19*, 1708–1713.

18. (a) Anderson, G. W.; Callahan, F. M. *J. Am. Chem. Soc.* **1960**, *82*, 3359–3363; (b) Valerio, R. M.; Alewood, P. F.; Johns, R. B. *Synthesis* **1988**, 786–789.
19. Liu, L.; Tanke, R. S.; Miller, M. J. *J. Org. Chem.* **1986**, *51*, 5332–5337.
20. Takeda, K.; Akiyama, A.; Nakamura, H.; Takizawa, S.; Mizuno, Y.; Takayanagi, H.; Harigaya, Y. *Synthesis* **1994**, 1063–1066.
21. Chevallet, P.; Garrouste, P.; Malawska, B.; Martinez, J. *Tetrahedron Lett.* **1993**, *34*, 7409–7412.
22. Loffet, A.; Galeotti, N.; Jouin, P.; Castro, B. *Tetrahedron Lett.* **1989**, *30*, 6859–6860.
23. Widmer, U. *Synthesis* **1983**, 135–136.
24. Dhaon, M. K.; Olsen, R. K.; Ramasamy, K. *J. Org. Chem.* **1982**, *47*, 1962–1965.
25. Ohta, S.; Shimabayashi, A.; Aona, M.; Okamoto, M. *Synthesis* **1982**, 833–834.
26. Shih, I.-L.; Chiu, L.-C.; Lai, C. T.; Liaw, W.-C.; Tai, D.-F. *Biotechnol. Lett.* **1997**, *19*, 857–859.
27. Galzigna, L. PCT Int. Appl. (1992), WO 9200320 A1 19920109; CAN 116:152418; AN 1992:152418.
28. A similar strategy to improve the solubility of GSH by preparing a lipophilic *S*-derivative prior to esterification was published while this manuscript was in preparation see Ref. [12b](#).
29. A related procedure produced a mixture of 54% *S*-palmitoyl and 46% *N*-palmitoyl glutathione: Vignais, P. V.; Zabin, I. *Biochim. Biophys. Acta* **1958**, *29*, 263–269.
30. Zhu, J.; Hu, X.; Dizin, E.; Pei, D. *J. Am. Chem. Soc.* **2003**, *125*, 13379–13381.
31. Zervas, L.; Photaki, I.; Ghelis, N. *J. Am. Chem. Soc.* **1963**, *85*, 1337–1341.